

Full Length Research Paper

Influence of some chemical compounds as antitranspirant agents on vase life of *Monstera deliciosa* leaves

Nermeen T. Shanan^{1*} and Emad A. Shalaby²

¹Department of Ornamental Horticulture, Faculty of Agriculture, Cairo University, Giza, Egypt 12613.

²Department of Biochemistry, Faculty of Agriculture, Cairo University, Giza, Egypt 12613.

Accepted 14 May, 2019

This study was designed to determine the influence of anti-transpiration agents, MgCO₃, Na₂CO₃ and glycerol, at four concentrations (2, 4, 6 and 8 %) on prolonging vase life of *Monstera deliciosa* cut leaves. For this purpose, two experiments were performed, through seasons 2008 and 2009. Each treatment sprayed three times 2, 4 and 6 days after cutting leaves. The results significantly revealed that glycerol at 2 or 4% extended vase life of *M. deliciosa* cut leaves by 7-folds of the control (7 days) and better than the other treatments. Also, glycerol treatment at the mentioned concentrations showed the lowest leaf weight reduction rate, as well as water loss rate, which obviously reflected on extending leaf vase life. The response of glycerol on prolonging leaf vase life was accompanied by a decrease in the degradation of pigments and protein as well as decrease the percentage of defense enzymes (superoxide dismutase and catalase) and this correlated with decreasing leaf water loss.

Key words: *Monstera deliciosa*, anti-transpirants vase life, enzyme.

INTRODUCTION

Demand for cut flowers and foliage plants for indoor decoration, is increasing dramatically around the world. The common potted foliage plants species include are the *Monstera deliciosa* and *Philodendron* sp. The foliage of these species are now also widely traded as cut floral greens (Will, 1985). Utilizing foliage plants for their attractive cut stems and the economic values. In general, the average vase life of *Monstera* is about 5 to 7 days and the globalization of the cut flower market means that *Monstera* greens have to be transported over long distances. Therefore, effective techniques are needed for preserving post-harvest quality in *Monstera* greens for a long time (Łukaszewska and Skutnik, 2003).

Anti-transpiration agents are grouped into three categories (Prakash and Ramachandran, 2000), firstly film-forming types (e.g. glycerol). Secondly, reflecting

materials which reflect the radiation falling on the upper surface of the leaves and thirdly stomatal closing types such as (MgCO₃ and Na₂CO₃) which affect the metabolic processes in leaf tissues (Ziv and Frederiksen, 1983; Osswald et al., 1984). Unfortunately, cut leaves have inefficient systems for regulating water loss, through transpiration and compensation through water uptake. The amount of water loss through transpiration is much higher leading to premature leaf wilting and death.. In cut leaves. The *Monstera* leaves have a large surface area, allowing for much water loss through transpiration (Geller and Smith, 1982).

To alleviate the damage from reactive oxygen species (ROS) such as superoxide anion (O₂⁻) and hydrogen peroxide (H₂O₂). Reactive oxygen species which cause lipid peroxidation and cell death (Mittler, 2002; Imlay, 2003), the foliage plants evolve enzymatic antioxidants such as superoxide dismutase (SOD) and catalase (CAT) (Xu et al., 2008). The objective of the present study was to evaluate the effect of different anti-transpiration

*Corresponding author: E-mail jnermeen_shanan@yahoo.com.

agents, on the foliage quality and vase life of *M. deliciosa*, through some morphological and chemical characters.

MATERIALS AND METHODS

Experimental design

This experiment was conducted using complete randomized design, during 2008 and 2009 seasons, in the laboratories of Ornamental Horticultural and Biochemistry Department. Mature, healthy and undamaged leaves of *M. deliciosa* plants were harvested in the morning, 1st of October, each year. Leaves were graded for uniformity. Each selected leaf was placed in glass container (750 ml) and filled with 500 ml of tap water. The three different treatments by anti-transpiration agents (MgCO₃, Na₂CO₃ and Glycerol) were used each at four concentrations (2, 4, 6 or 8% w/v). Each treatment contained three replicates, each represented by three leaves. The treatments have individual control (spraying with tap water). However, the average of three controls was calculated and used in the statistical analysis. Every treatment sprayed three times, i.e. 2, 4, 6 days. The experiment started with 500 ml as volume solution of all used anti-transpiration treatments.

All the containers were placed under laboratory controlled environmental conditions; temperature at 23±1°C, relative humidity 60% and 1500 Lux of continuous light (10 to 14 h day/night). The data were taken 2 days intervals till the end of the experiment. The characters studied were vase life, leaf water loss and reduction rate of leaf weight.

Pigments determination

Pigments was determined as chlorophyll and carotenoid contents using the method of Holden (1965).

Electrophoretic fractionation of soluble proteins

Polyacrylamide gel electrophoreses in the presence of sodium dodecyl sulphate (SDS-PAGE), was used for determining the molecular weight of protein fractions (Total soluble protein) according to method of Laemmli (1970). Standard molecular weight proteins marker was obtained from Sigma, this marker contained different proteins molecular weights, i.e. 250, 150, 100, 70, 50, 40, 30, 20, 15, 10 and 5 kDa.

Enzyme extracts

A ground sample (1.0 g) was homogenized in 3 mL of 50 mM phosphate buffer pH 7.0, 1% PVP, 1 mM ascorbate at 4°C. After centrifugation at 15,000×g for 15 min, the supernatant was collected according to Vitória et al. (2001).

Catalase (CAT; EC 1.11.1.6)

Catalase activity was determined as H₂O₂ consumption measured by the decrease in absorbance at 240 nm according to the method of Aebi (1983). The assay buffer contained 50 mM KH₂PO₄/K₂HPO₄ (pH 7.0), 10 mM H₂O₂. Extinction coefficient of 39.4 mM⁻¹cm⁻³ was used to calculate activity. Enzyme activity was expressed in μM H₂O₂ hydrolyzed per min.

Superoxide dismutase (SOD; EC 1.15.1.1)

Superoxide dismutase activity was measured by the photochemical method as described by Beauchamp and Fridovich (1971). One unit of SOD activity was defined as the amount of enzyme required to cause 50% inhibition of the rate of nitro blue tetrazolium (NBT) reduction at 560 nm in the presence of riboflavin and light. The reaction mixture contained 45 mM potassium phosphate buffer, pH 7.0, containing 0.1 mM EDTA and 13 mM methionine, 0.17 mM NBT in ethanol, 0.007 mM riboflavin and enzyme aliquot. Blanks were kept in the dark whereas the treatments.

Statistical analysis

Data were subjected to analysis of variance, means were compared using the "Least Significant Difference test (New LSD) at 0.01 and 0.05 levels, using M-STATE software package (1987).

RESULTS

Vase life

Data in Table 1 showed that in both seasons, all foliar applications significantly increased the vase life of treated leaves, as compared to the control (Figure 1). The cut leaves treated with glycerol at 4 or 2 % has the highest values of vase life among all the other treatments.

Leaf water loss

Anti-transpirants have been proposed as a method to reduce water loss, and enhance the water status of plants. Data in Table 2 showed that the effect of using foliar treatments by glycerol, MgCO₃ and Na₂CO₃ at different concentrations, decreased the rate of leaf water loss as compared to control. The best results were obtained from glycerol treatment either at 2 or 4% in both seasons and sodium carbonate at 4% and 8% in the first and second seasons, respectively comparing with control.

The vase life of leaves treated by glycerol at 4% was 7 folds that of control, and this was accompanied in treated leaves by the rate of water loss.

Reduction rate of leaf weight

Data in Table 2 represented the reduction rate in leaf weight (g.day⁻¹) as affected by anti-transpiration agents. In both seasons, anti-transpiration treatments decrease significantly the reduction rate of leaf weight comparing with the control. Glycerol treatment at concentration 2 or 4% had the lowest value of leaf weight reduction rate which obviously reflected on leaf vase life.

Total pigments

It is obvious from Table 3 that, the total chlorophyll percentage after 6 days was higher with any concentrations

Table 1. Effect of anti-transpiration agents at different concentrations on vase life of *Monstera deliciosa* in both seasons.

| Anti-transpiration agents | Concentrations (%) | First season | Second season |
|---------------------------------|--------------------|------------------|---------------|
| | | Vase life (days) | |
| Control | | 6.33 ±0.33 e | 7.00 ±0.76 e |
| Na ₂ CO ₃ | 2 | 11.00 ±1.64 cd | 19.33±2.46 c |
| | 4 | 12.33 ±1.74 cd | 14.00±1.82 cd |
| | 6 | 13.33 ±1.80 cd | 10.33±1.49 cd |
| | 8 | 14.00 ±1.36 cd | 18.17±2.17 c |
| MgCO ₃ | 2 | 15.00 ±1.21 cd | 11.33±1.02 cd |
| | 4 | 15.67 ±1.00 cd | 18.50±1.78 c |
| | 6 | 16.33 ±1.91 b-d | 14.00±2.14 cd |
| | 8 | 17.00 ±2.02 b-d | 26.67±2.92 b |
| Glycerol | 2 | 44.00 ±3.09 a | 57.33 ±3.67 a |
| | 4 | 46.00 ±3.95 a | 54.00 ±3.24 a |
| | 6 | 21.00 ±2.44 bd | 18.83±1.68 c |
| | 8 | 20.00 ±1.68 bd | 26.67±2.06 b |

Values are the mean of three replication±SD. Mean separation among treatments was done by New LSD test (0.05); Mean values followed by different letters are significantly different.

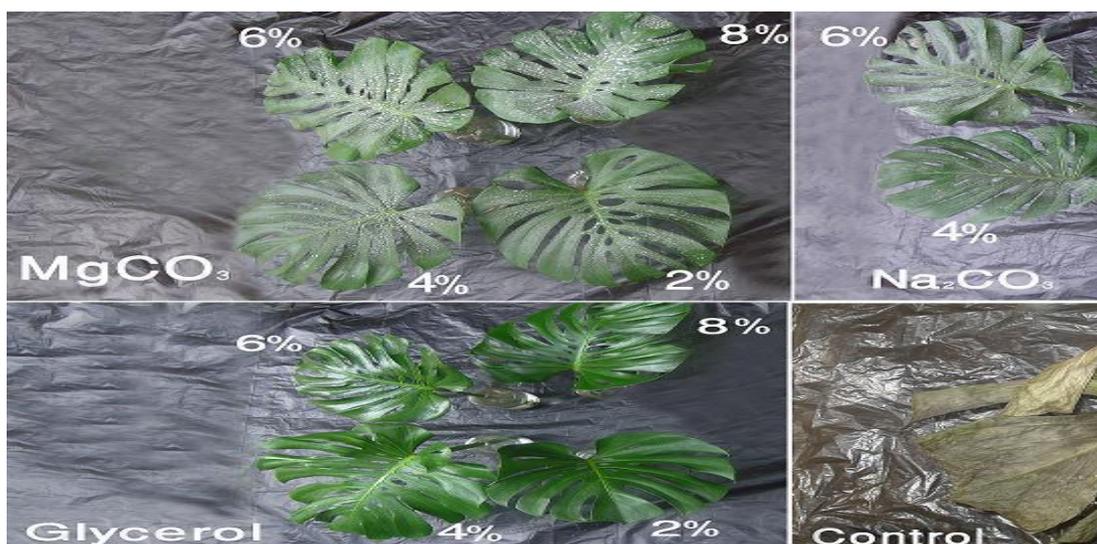


Figure 1. The effect of different antitranspiration agents concentration on the vase life of *Monstera deliciosa* (after 6 days).

of the used antitranspiration agents than control. The highest values of total chlorophyll were with MgCO₃ at 4 and 2% among all other treatments. The nearest highest value to that after 2 days was with 2% glycerol. While the total carotenoids was the highest with the three concentrations of glycerol (2, 6 and 8%) and with MgCO₃ at 6 and 8% after 2 days. Contrary, after 6 days, most of the concentrations of the used antitranspiration do not influence the total carotenoids expect the minute increase with 2% Na₂ CO₃ and 8% glycerol that the pigments

concentrations were decreased after 6 days in all treatments

Enzyme activities

The activities of Catalase and Superoxide dismutase (CAT and SOD) enzymes were significantly (P<0.01) stimulated, and this stimulation reached its maximum at untreated plant (control) after 6 days for both enzymes. Enzyme activities were significantly stimulated and

Table 2. Effect of anti-transpiration agents at different concentrations on the rate of leaf water loss and reduction rate of leaf weight of *Monstera deliciosa* during the first and second seasons.

| Antitranspiration agents | Concentrations (%) | First season | | Second season | |
|---------------------------------|--------------------|--|--|--|--|
| | | Leaf water loss (g.dm ⁻² .day ⁻¹) | Reduction rate of leaf weight (g.day ⁻¹) | Leaf water loss (g.dm ⁻² .day ⁻¹) | Reduction rate of leaf weight (g.day ⁻¹) |
| Control | | 0.32±0.03 a | 0.40±0.04 a | 0.30 ±0.01 a | 0.53± 0.02 a |
| MgCO ₃ | 2 | 0.30 ±0.03 a | 0.12±0.01 de | 0.20 ±0.02 cd | 0.12±0.01 efg |
| | 4 | 0.25 ±0.03 bc | 0.14±0.02 cde | 0.19 ±0.02 cd | 0.14 ±0.01 ef |
| | 6 | 0.21 ±0.03 cd | 0.17±0.02 bcd | 0.27 ±0.03 b | 0.23 ±0.03 c |
| | 8 | 0.20 ±0.03 cd | 0.24±0.04 b | 0.15 ±0.02 e | 0.34 ±0.05 b |
| Na ₂ CO ₃ | 2 | 0.14 ±0.02 f | 0.16±0.02 cd | 0.22 ±0.02 c | 0.10 ±0.01 efg |
| | 4 | 0.09 ±0.01g | 0.42 ±0.05 a | 0.14 ±0.02 e | 0.30 ±0.04 c |
| | 6 | 0.17 ±0.02 cd | 0.15±0.02 cde | 0.15 ±0.02 e | 0.18 ±0.02 cd |
| | 8 | 0.14 ±0.01 d | 0.10±0.01 ef | 0.09 ±0.01 g | 0.12 ±0.01 fg |
| Glycerol | 2 | 0.07 ±0.01 e | 0.07±0.01 fg | 0.06 ±0.01 h | 0.09±0.02 g |
| | 4 | 0.05 ±0.01 e | 0.06± 0.01g | 0.06 ±0.01 h | 0.08±0.01 g |
| | 6 | 0.14 ±0.01 d | 0.12 ±0.01 ef | 0.15 ±0.01 e | 0.15±0.01 def |
| | 8 | 0.14 ±0.01 d | 0.18 ±0.02 bc | 0.12 ±0.02 f | 0.20 ±0.03 cd |

Values are the mean of three replication±SD. Mean separation among treatments was done by New LSD test (0.05). Mean values followed by different letters are significantly different.

Table 3. The pigments percentage (%) after 2 and 6 days from treatments leaves of *Monstera deliciosa* with anti-transpiration agents with three concentrations.

| Antitranspiration agents | Concentration (%) | Total chlorophyll | Total carotenoids | Total chlorophyll | Total carotenoids |
|---------------------------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| | | After 2 days | | After 6 days | |
| Control | | 1.13±0.10 h | 0.33±0.00 e | 0.96±0.15 l | 0.30±0.00 b |
| MgCO ₃ | 2 | 0.99±0.00 k | 0.2±0.00 j | 1.72±0.15 b | 0.24±0.00 g |
| | 4 | 1.01±0.20 j | 0.21±0.05 j | 1.96±0.23 a | 0.21±0.10 h |
| | 6 | 1.27±0.10 f | 0.39±0.00 c | 1.38±0.10 k | 0.25±0.05 f |
| | 8 | 1.51±0.30 c | 0.42±0.10 b | 1.68±0.30 d | 0.24±0.00 g |
| Na ₂ CO ₃ | 2 | 1.23±0.00 g | 0.31±0.00 f | 1.42±0.00 j | 0.31±0.05 a |
| | 4 | 1.3±0.15 e | 0.24±0.00 h | 1.69±0.15 c | 0.24±0.00 g |
| | 6 | 1.38±0.00 d | 0.29±0.00 g | 1.55±0.15 h | 0.25±0.00 f |
| | 8 | 1.38±0.12 d | 0.21±0.05 j | 1.63±0.20 e | 0.25±0.00 f |
| Glycerol | 2 | 1.71±0.13 a | 0.48±0.00 a | 1.56±0.15 g | 0.28±0.03 d |
| | 4 | 1.06±0.10 i | 0.23±0.02 i | 1.58±0.15 f | 0.26±0.00 e |
| | 6 | 1.68±0.30 b | 0.35±0.10 d | 1.58±0.20 f | 0.29±0.04 c |
| | 8 | 1.68±0.20 b | 0.41±0.10 b | 1.54±0.10 i | 0.31±0.02 a |

Values are the mean of three replication±SD. Mean separation among treatments was done by New LSD test (0.01). Mean values followed by different letters are significantly different.

Table 4. Effect of treating *Monstera deliciosa* leaves with anti-transpiration agent on superoxide dismutase (SOD) and catalase (CAT) activities of (After 6 days).

| Anti-transpiration agents | Concentration (%) | SOD (Unit/mgProtein) | CAT ($\mu\text{mol/mgprotein/min}$) |
|---------------------------------|-------------------|----------------------|---------------------------------------|
| Control | | 7.40 \pm 0.20 a | 2.90 \pm 0.00 a |
| MgCO ₃ | 2 | 7.10 \pm 0.30 c | 2.76 \pm 0.00 b |
| | 4 | 7.20 \pm 0.10 b | 2.70 \pm 0.00 c |
| | 6 | 6.70 \pm 0.50 d | 2.50 \pm 0.20 g |
| | 8 | 6.65 \pm 0.40 e | 2.53 \pm 0.00 f |
| Na ₂ CO ₃ | 2 | 6.32 \pm 0.15 f | 2.65 \pm 0.20 d |
| | 4 | 5.50 \pm 0.00 j | 2.57 \pm 0.30 e |
| | 6 | 5.40 \pm 0.20 k | 2.50 \pm 0.00 g |
| | 8 | 5.00 \pm 0.00 l | 2.32 \pm 0.10 i |
| Glycerol | 2 | 5.90 \pm 0.20 i | 2.40 \pm 0.10 h |
| | 4 | 6.00 \pm 0.20 h | 2.52 \pm 0.10 fg |
| | 6 | 6.30 \pm 0.10 f | 2.70 \pm 0.00 c |
| | 8 | 6.20 \pm 0.10 g | 2.76 \pm 0.00 b |

Values are the mean of three replication \pm SD. Mean separation among treatments was done by New LSD test (0.01). Mean values followed by different letters are significantly different.

negativity correlated, in most cases, with the levels of antitranspirants agents treatment except with glycerol 2 and 4% (Table 4).

Fractionation of total soluble protein

Treatment with anti-transpiration agents not only affected vase life, pigments content but also, the leaf chemical constituents, that is, soluble proteins. Data recorded in Table 5 showed that, the number of protein bands changed from treated and untreated leaves (ranged from 3 bands in plant treated with 2% sodium carbonate to 13 bands in leaves treated with 6% glycerol after 6 days) when compared to untreated leaf (8 bands). From Table 5 the results indicated that, no protein bands of high molecular weights (100-300 kDa) and lower molecular weights (10-0.01 kDa) were detected in untreated leaves, but these bands appear after treatment with the agents at different concentrations.

The results also indicate that the tested anti-transpiration agents affect on physiological functions of treated leaves through synthesis of different short proteins these new proteins might help leaves for defending against stress by stabilizing the quaternary structure of proteins in the membrane as well as the related enzymes, which in turn reflected on extended vase life.

DISCUSSION

Foliar applications significantly increased the vase life of treated leaves. Similarly, Ponce et al. (2009) found that

glycerol or sorbitol (1%) extended the shelf-life of fresh apples in 10 days. De-Stigter (1981) treated the cut flowers with the commercial preservatives (8-HQC + 2% Suc) to diminish transpirational loss and maintain flower turgidity and therefore extended their vase life (Łukaszewska and Skutnik, 2003). However, some treatments decreased the rate of leaf water loss, these findings in agreement with Jones et al. (2004) who found that, anti-transpiration treatments did not decrease solution uptake by the holly stems, leading improve marketability of branches. Using anti-transpirants improved the water use efficiency and reduced leaf transpiration rate by 87 to 93% (Nasraoui, 1993; Bora and Mathur, 1998; Makus, 1997). Francisco and Rubio (2009) found that the Anti-transpirant (Pinolene) significantly reduced water uptake but no effect was found with control solution. Moreover, Shen et al. (1999) and Yancey et al. (2005) found that, glycerol can function either as an osmolyte, contributing to the maintenance of water balance, or as an osmoprotectant, allowing the operation of many cellular processes during osmotic stress. Thus, this result supports the present findings, concerning the effect of the glycerol on decreasing the water loss which could enhance the vase life. In addition, it was noticed a parallel increase in the rate of the water loss with increasing both MgCO₃ and glycerol over 6%, which were inversely related to vase life. Dubois and Joyce (1992) found the same result in ornamental plants. Generally, it could be concluded that glycerol can be used as a tool for reducing plant water loss, which could be resulted from closing stomata openings and reducing the transpiration rate; as mentioned by previous

Table 5. SDS-Electrophoresis analysis of soluble proteins produced by treatment of *Monstera deliciosa* with different concentration of antitranspirants agents (after 6 days).

| Protein band | Molecular weight (kDa) | Control | Anti-transpiration agents | | | | | | | | | | | |
|--------------|------------------------|---------|---------------------------|------|------|------|---------------------------------|------|------|------|-------------------|------|------|------|
| | | | Glycerol | | | | Na ₂ CO ₃ | | | | MgCO ₃ | | | |
| | | | Concentrations (%) | | | | | | | | | | | |
| | | | 2 | 4 | 6 | 8 | 2 | 4 | 6 | 8 | 2 | 4 | 6 | 8 |
| 1 | 292.00 | - | - | - | - | - | - | 4.0 | - | - | - | - | - | 8.0 |
| 2 | 277.37 | - | - | - | - | - | - | - | - | - | - | - | - | 43.0 |
| 3 | 208.00 | - | - | - | 5.0 | - | - | - | - | - | - | - | - | 3.9 |
| 4 | 187.00 | - | - | - | - | - | - | 4.2 | - | - | - | - | - | - |
| 5 | 169.33 | -- | -- | -- | - | -- | -- | -- | -- | 5.0 | -- | - | -- | - |
| 6 | 138.00 | - | - | - | 5.0 | - | - | - | - | - | - | -- | - | - |
| 7 | 118.00 | - | - | - | 7.5 | 15.2 | - | - | - | - | - | - | - | 4.3 |
| 8 | 110.00 | - | - | - | 7.5 | - | 42.0 | - | - | - | - | - | - | - |
| 9 | 98.00 | 16.1 | - | - | 5.0 | - | - | -- | - | - | - | 7.5 | - | - |
| 10 | 96.00 | - | - | - | - | - | - | - | - | - | 15.0 | - | - | - |
| 11 | 94.00 | 7.5 | - | - | - | 10.0 | - | - | - | - | - | - | - | - |
| 12 | 90.50 | - | - | - | - | 5.0 | - | - | - | 7.5 | - | - | - | -- |
| 13 | 87.50 | 7.6 | - | 17.1 | - | - | -- | - | -- | - | - | - | -- | 5.4 |
| 14 | 84.26 | - | - | - | 10.9 | 7.6 | - | - | - | 5.0 | - | -- | - | 3.4 |
| 15 | 76.62 | - | -- | - | 11.0 | - | - | 10.1 | - | 5.0 | -- | - | - | - |
| 16 | 74.8 | 16.1 | - | - | - | - | - | - | - | - | - | - | - | - |
| 17 | 71.34 | 17.5 | - | - | - | - | - | - | 16.1 | - | - | - | - | 3.5 |
| 18 | 66.93 | - | - | -- | - | - | - | - | 7.5 | - | - | - | - | - |
| 19 | 50.00 | 7.5 | - | - | - | - | - | 5.5 | - | - | - | - | - | - |
| 20 | 44.72 | - | - | - | 9.0 | - | - | - | - | -- | - | - | - | - |
| 21 | 40.00 | - | - | - | - | - | - | - | 15.0 | - | - | - | -- | - |
| 22 | 36.34 | - | - | 18.1 | - | - | - | - | - | - | 15.0 | - | 15.0 | -- |
| 23 | 19.82 | - | -- | - | - | - | - | - | 25.1 | - | - | 7.5 | - | - |
| 24 | 18.97 | -- | - | - | - | -- | -- | 5.6 | - | 5.6 | - | - | - | - |
| 25 | 16.61 | 16.0 | 20.0 | - | 7.5 | - | - | 10.2 | 15.0 | - | - | 27.2 | 7.6 | - |
| 26 | 10.2 | 13.2 | - | 30.1 | 5.0 | - | - | 5.5 | 7.8 | - | 15.0 | - | 20.0 | - |
| 27 | 7.77 | - | - | - | - | 10.1 | - | - | 7.5 | 7.4 | - | - | 35.0 | - |
| 28 | 6.04 | - | - | - | - | - | - | - | - | - | - | - | - | - |
| 29 | 4.14 | - | - | 20.1 | - | - | - | - | - | - | - | - | - | - |
| 30 | 1.51 | - | - | 15.0 | 5.0 | - | - | 15.1 | - | 7.9 | - | 27.5 | - | - |
| 31 | 0.55 | -- | 50.0 | - | 7.5 | 7.5 | 19.8 | 14.2 | - | 7.8 | 28.0 | - | - | 4.4 |
| 32 | 0.01 | - | 19.6 | - | 20.1 | - | 20.0 | 15.1 | 7.3 | 50.2 | - | - | 30.0 | 22.0 |
| No. of bands | | 8.0 | 3.0 | 5.0 | 13.0 | 6.0 | 3.0 | 10.0 | 8.0 | 9.0 | 4.0 | 4.0 | 5.0 | 9.0 |

investigators and leading to increased leaf vase life. Anti-transpiration treatments decrease significantly the reduction rate of leaf weight; it could state that a treatment which showed a slight rate of water loss could has a slight reduction rate of leaf weight which might be reflected on decreasing of transpiration rate. These results are supported by Abd El-kader et al. (2006) who found that foliar sprays of magnesium carbonate as antitranspirants on 'Williams banana' increased growth parameters. The same trend was found by Mofteh and Al-Humaid (2006) on tuberose plants and Liang et al. (2002) who reported that water consumption was less for the anti-transpiration treated plants.

Pigments percentage (chlorophyll and carotenoids) were increased by all treatments, these results were correlated with the vase life results, however, high concentration of pigments in treated leaves could be due to the effect of anti-transpirant, in improving water use efficiency, by reducing leaf transpiration rate via increasing leaf reflecting or inducing stomata closure. These results are in agreement with those obtained by Rabiza-Świder and Skutnik (2004) who found that, postharvest longevity of *Zantedeschia* and *Hosta* was extended by inhibiting leaf senescence through delaying chlorophyll loss and soluble protein degradation. Nevertheless, Abd El-Kader et al. (2007) recorded that

spraying antitranspirants increased growth parameters. El-Abd (1996) on citrus, Ranney et al. (1989) on cherry trees recorded that pruning and antitranspirant were successful in delaying plant water stress, and relative growth rate.

Enzyme induction could be due to the effect of antitranspirant agent in improving water use efficiency, by reducing leaf transpiration rate and decrease leaf water loss. This may be due to increasing water loss value in control, than treated leaves and led to increase the free radicals formation (O_2^- , H_2O_2 and OH^-) this correlated with increasing various defense enzymes especially antioxidant enzymes (Catalase and superoxide dismutase). Nearly, same results were obtained by Zwiazek and Blake (1990), who observed that, drought caused a reduction in sterols, phospholipids, and sterol/phospholipid ratio, along with the increase in membrane leakage in dehydrating black spruce. A shift in phospholipid concentration could explain the membrane damage by induced peroxidation of lipids. This is resulted from the formation of free radicals (O_2^- , H_2O_2 , and/or OH^-), which destabilize chloroplast, mitochondrial, and/or microsomal membranes. Another study reported that higher plants have active oxygen-scavenging systems consisting of several antioxidant enzymes, such as ascorbate peroxidase (APX), guaiacol peroxidase (GPX) and catalase (CAT). These systems protect membranes from the deleterious effects of ROS, such as superoxide radicals, hydrogen peroxide (H_2O_2), hydroxyl radicals and singlet oxygen, which are produced at elevated rates when plants are exposed to abiotic stress conditions (Noctor and Foyer 1998). The superoxide radical is dissimulated to H_2O_2 by SOD, CAT and APX metabolize H_2O_2 into H_2O . Mohammadkhani and Heidari (2007) found that maize, under drought stress; the activities of GPX, APX and CAT were increased in roots and shoots.

Anti-transpiration agents affected the leaf chemical constituents, that is, soluble proteins. No protein bands of high and lower molecular weights were detected in untreated leaves, but appear only after treatment with the three used agents at different concentrations. These results may be due to the effect of different anti-transpiration treatment on induction or inhibition or both on gene expression which led to absence, presence or increase/ decrease of protein bands intensity. These results could be explained by Taravati et al. (2007) who mentioned that, hydroxyl groups in polyols are thought to form a hydration sphere around macromolecules, thus protecting cells against stress by stabilizing the quaternary structure of proteins such as membranes and enzymes.

Conclusion

It could be concluded that, glycerol at concentration is either 2 or 4% exceeded the vase life leaf about 7-folds, and this was accompanied by lowering the reduction rate

leaf water loss and leaf weight as compared to other treatments. In addition, the effect of glycerol at the mentioned concentrations could decrease the degradation of pigments and proteins and increase the percentage of defense enzymes (SOD and CAT) and this correlated with its ability to decrease leaf water loss.

REFERENCES

- Aebi H (1983). Catalase, in H Bergmeyer (Ed.) Methods of Enzymatic Analysis. Verlag Chemie, Weinheim, Adamse, 3: 273-277.
- Abd El – kader AM, Saleh M MS, Ali MA (2006). Effect of soil moisture levels and some antitranspirants on vegetative growth, leaf mineral content, yield and fruit quality of Williams's banana plants. J. Appl. Sci. Res., 2(12): 1248-1255.
- Beauchamp C, Fridovich I (1971). Superoxide dismutase improved assays and an assay applicable to acrylamide gels. Anal. Biochem., 44: 276-287.
- Bora KK, Mathur SR (1998). Some plant growth regulator as antitranspirants in soybean. Ann. Plant Physiol., 12: 175-177.
- De-Stigter HCM (1981). Water balance aspects of cut and intact 'Sonia' rose petals and effect of glucose, 8HQS and aluminum sulfate. Acta Hortic., 113: 97-107.
- Dubois P, Joyce D (1992). Preservation of fresh cut ornamental plant material with glycerol. Postharvest Biol. Technol., 2: 145-153.
- El-Abd AAA (1996). Studied on the effect of drainage water and/or antitranspirants on growth and some chemical constituents of some Citrus rootstock. M.Sc.Thesis, Fac. of Agric.Kafr El- Shikh, Tanta Univ. Egypt.
- Francisco MA, Rubio JS (2009). Effects of Antitranspirant Spray and Potassium: Calcium: Magnesium Ratio on Photosynthesis, Nutrient and Water Uptake, Growth, and Yield of Sweet Pepper. J. Plant Nutr., 32(1): 97–111.
- Geller GN, Smith WK (1982). Influence of leaf size, and arrangement on temperature and transpiration in three high-elevation, large-leaved herbs. Oecologia (Berl), 53: 227-234.
- Holden M (1965). Chlorophyll, In chemistry and biochemistry of plant pigments. Ed. Goodwin, T. W., Academic Press, London, pp. 462-88.
- Imlay JA (2003). Pathways of oxidative damage. Annu. Rev. Microbiol., 57: 395–418.
- Jones LM, Cochran K, Anderson GA, Ferree CD (2004). Effect of preservatives and cold storage on postharvest performance of deciduous holly branches. Hortechonol., 14(2): 230-234.
- Laemmli UK (1970). Cleavage of structural proteins during the assembly of head of bacteriophage T4. Nature (London), 227: 680-685.
- Liang Z, Zhang F, Shao M, Zhang J (2002). The relations of stomata conductance, water consumption, growth rate to leaf water potential during soil drying and rewatering cycle of wheat (*Triticum aestivum*). Bot. Bull. Acad. Sin., 43: 187-192.
- Łukaszewska A, Skutnik E (2003). Guide to florists. Copyright by, p. 156.
- Makus DJ (1997). Effect of an antitranspirant on cotton grown under conventional tillage systems. Proc. betwide cotton conf., New Orleans, LA, USA. January, 6(10): 642-644.
- MASTATE Version4. (1987). Software program for the design and analysis of agronomic research experiments. Michigan State Univ., MS., U.S.A.
- Mittler R (2002). Oxidative stress, antioxidants and stress tolerance. Trends Plant Sci., 7: 405–410.
- Moftah AE, Al-Humaid AI (2006). Response of vegetative and reproductive parameters of water stressed Tuberose plants to vapor grad and Kaolin Antitranspirants. J. King Saud Univ. Agric. Sci., 18(2): 127-139.
- Mohammadkhani N, Heidari R (2007). Water stress induced by polyethylene glycol 6000 and sodium chloride in two maize cultivars. Pak. J. Biol. Sci., 11(1): 92-97.
- Nasraoui B (1993). Role of antitranspirant films in protecting plants against fungal diseases. Annals of I, Natl. Inst. Agron. Res. Tunisia,

- 66: 125-135.
- Noctor G, Foyer CH (1998). Ascorbat and glutathione: Keeping active oxygen under control. *Ann. Rev. Plant Physiol. Plant Mol. Biol.*, 49: 249-279.
- Osswald WM, Neihuss M, Huber W, Elstner EF (1984). Support of non-host resistance by artificial leaf coating. *Z. Krankh plants. Plant protect.*, 91: 337-341.
- Ponce GL, Carbonari R, Ademar BL (2009). Active Packaging using ethylene absorber to extend shelf- life, Rio de Janeiro, RJ, Brazil. INAC.
- Prakash M, Ramachandran K (2000). Effect of chemical Ameliorants in Brinjal (*Solanum melongena* L.) under moisture strees conditions. *J. Agron. Crop Sci.*, 185: 237-239.
- Rabiza-Swider J, Skutnik E (2004). Effect of light on senescence of cut leaves of *Zantedeschia aethiopica* Spr. and *Hosta* Tratt. '*Undulata Erromena*'. *Folia Hortic.*, 16(1): 161-166.
- Ranney TG, Bassuk NL, Whitlow TH (1989). Effect of transplanting practice on growth and water relation of "colt" cherry trees during reestblishment. Department of Horticulture, Aalabama Agricultural Experimental Station, Auburn University, Auburn, AL 36849, USA.
- Shen B, Hohmann S, Jensen RG, Bohnert HJ (1999). Roles of Sugar Alcohols in Osmotic Stress Adaptation. Replacement of Glycerol by Mannitol and Sorbitol in Yeast. *Plant Physiol.*, 121: 45-52.
- Taravati A, Shokrzadeh M, Ebadi AG, Valipour P, Molla TA, Farrokhi F (2007). Various effects of sugar and polyols on the protein structure and function: Role as osmolyte on protein stability. *World Appl. Sci. J.*, 2(4): 353-365.
- Vitória AP, Lea PJ, Azevedo RA (2001). Antioxidant enzymes responses to cadmium in radish tissues. *Phytochemistry*, 57: 701-710.
- Will AA (1985). Foliage for flower arrangements from your Florida garden. *Proc. Fla. State Hortic. Soc.*, 98: 349-350.
- Xu PL, Guo YK, Bai JG, Shang L, Wang XJ (2008). Effects of long-term chilling on ultrastructure and antioxidant activity in leaves of two cucumber cultivars under low light. *Plant Physiol.*, 132: 467-478.
- Yancey, PH (2005). Organic osmolytes as compatible, metabolic and counteracting cytoprotectants in high osmolarity and other stresses. *J. Exp. Biol.*, 208: 2819-2830.
- Ziv O, Frederiksen RA (1983). Control of foliar disease with epidermal coating materials. *Plant Dis.*, 67: 212-214.
- Zwiazek TJ, Blake TJ (1990). Effects of pre-conditioning on electrolyte leakage and lipid composition in black spruce (*Picea mariana*) stresses with polyethylene glycol. *Plant Physiol.*, 79: 71-77.