

International Journal of Animal Breeding and Genetics ISSN 2756-3650, Vol. 14 (4), pp. 001-005, April, 2025. Available online at www.internationalscholarsjournals.org © International Scholars Journals

Author(s) retain the copyright of this article.

Full Length Research Paper

# Impact of Biological Feed Additives on Broiler Chicken Growth Performance

A. Ashayerizadeh<sup>1\*</sup>, N. Dabiri<sup>1,2</sup>, Kh. Mirzadeh<sup>1</sup> and M. R. Ghorbani<sup>1</sup>

<sup>1</sup>Department of Animal Science, Ramin Agricultural and Natural Resources University, Ahvaz, Iran. <sup>2</sup>Faculty of Agricultural, Animal Science Department, Islamic Azad University, Karaj Branch, Karaj, Iran.

# Accepted 22 March, 2025

The present study aimed at evaluating the effect of the probiotic Primalac, prebiotic Biolex-MB, a combination of the two supplements (probiotic plus prebiotic) and one growth promoter antibiotic (Flavomycin) on performance results of broiler chickens. Three hundred day old Ross 308 broilers were equally distributed into 30 floor pens and reared for 42 day. A basal diet was also supplemented with Flavomycin (650 g/ton<sup>-1</sup>), Primalac (900 g/ton<sup>-1</sup>), Biolex-MB (2000 g/ton<sup>-1</sup>) and mixture of Primalac plus Biolex-MB (synbiotic) for starter (1 to 21 days) and grower (22 to 42 days) periods, resulting 5 dietary treatments were prepared including control group. Each dietary treatment was fed ad-libitum to six replicate group of 10 birds at the beginning of rearing period. Birds and feed were weighed weekly. The daily body weight gain and feed efficiency were significantly (P<0.05) more improved by the synbiotic treatments compared with the control broilers. The birds under antibiotic treatment had higher breast and thigh percent as compared to control birds (p<0.05). Also, supplementation of broiler's diet with prebiotic significantly (p<0.05) increased gizzard percent. The results of present study showed that probiotic and prebiotic can be used as non-antibiotic growth promoter feed additives to improve broiler chickens growth indices.

Key words: Antibiotic, prebiotic, probiotic, performance.

# INTRODUCTION

Antimicrobials have been used as feed supplement for more than 50 years in poultry feed to enhance the growth performance and to prevent diseases in poultry. However, in recent years great concern has arisen about the use of antibiotics as supplement at sub-therapeutic level in poultry feed due to emergence of multiple drug resistant bacteria (Wray and Davies, 2000). As a conse-quence, it has become necessary to develop alternatives using either beneficial microorgansims or nondigestible ingredients that enhance microbial growth. A probiotic is a culture of a single bacteria strain, or mixture of different strains, that can be fed to an animal to improve some aspect of its health (Griggs and Jacob, 2005). Probiotics are also referred to as direct fed microbials (DFM). On the other hand, a prebiotic was defined as nondigestible

food ingredient that beneficially affects the host,

selectively stimulating the growth or activity, or both, of one or a limited number of bacteria in the colon (Gibson and Roberfroid, 1995).

The efficacy of probiotics may be potentiated by several methods: the selection of more efficient strains, gene manipulation, the combination of several strains, and the combination of probiotics and synergistically acting components. This approach seems to be the best way of potentiating the efficacy of probiotics and is widely used in practice. A way of potentiating the efficacy of probiotic preparations may be the combination of both probiotics and prebiotics as synbiotics, which may be defined as a mixture of probiotics and prebiotics that beneficially affects the host by improving the survival and implantation of live microbial dietary supplements in the gastrointestinal (GI) tract. Those effects are due to activating the metabolism of one or a limited number of health-promoting bacteria or by selectively stimulating their growth, which improved the welfare of the host, or both (Gibson and Roberfroid, 1995). Lactobacilli and enterococci are among the wide variety of microbial species that have been used extensively as probiotics

<sup>\*</sup>Corresponding author. E-mail: amin.ashayerizadeh@yahoo.com. Tel: +98-9173099064. Fax: +98-7297262150.

(Patterson and Burkholder, 2003). Primalac is a kind of commercial probiotic that contains at least  $1 \times 10^8$  CFU g<sup>-1</sup> *Lactobacillus casei, Lactobacillus acidophilus, Bifidobacterium thermophilum,* and *Enterococcus faesium* (Chichlowski et al., 2007a, b). After feeding of probiotics, improvements in growth performance and feed efficiency have been reported in broiler chickens (Cavazzoni et al., 1998; Zulkifli et al., 2000; Kabir et al., 2004; Samli et al., 2007). Two of the most commonly

oligosaccharides studied prebiotic are fructooligosaccharides (FOS) and mannanoligosaccharides (MOS). FOS can be found naturally in some cereal crops and onions (Bailey et al., 1991). MOS is obtained from the cell wall of veast (Saccharomyces cerevisiae). Biolex-MB is a commercial prebiotic of the mannanoligosaccharides family, which is obtained by extraction from the outer cell wall of the yeast Saccharomyces cerviciae. Several typical probiotics contain either of these oligosaccharides, thereby comprising a synbiotic. The combination of a probiotic and prebiotic in one product has been shown to confer benefits beyond those of either on its own (Gallaher and Khil, 1999).

The search for new additives effective on animal growth and free from harmful side effects on consumers health is still continuing. The aim of this study was comparing the effects of the antibiotic flavomycin and primalac and Biolex-MB as alternatives for the growth stimulating antibiotics on growth performance and carcass characteristics of broiler chickens.

# MATERIALS AND METHODS

# Birds and housing

Three hundred 1 day old broiler chicks (Ross 308) were obtained from a commercial hatchery. The birds were randomly divided into 5 groups (60 birds/group) and housed in pens of identical size (120  $\times$ 100 cm) in a deep litter system with a wood shaving floor. Each group had 6 replicates (10 birds/pen). The birds had free access to water and feed. Environmental temperature in the first week of life was 34°C and decreased to 20°C until the end of the experiment.

# Diets

A basal diet was formulated and considered as control according to recommendation of NRC, 1994 for starter (1 to 21 days), and grower (22 to 42 days) diets. Four tested diets were formulated by supplemented the basal control diet with antibiotic Flavomycin (650 g/ton<sup>-1</sup>), probiotic (Primalac, 900 g/ton<sup>-1</sup>), prebiotic (Biolex-MB, 2000 g/ton<sup>-1</sup>), and mixture of Primalac (900 g/ton<sup>-1</sup>) plus Biolex-MB (2000 g/ton<sup>-1</sup>) as synbiotic, respectively. The composition of the diets are shown in Table 1.

#### Growth performance traits

All birds were weighed individually after their arrival from the hatchery to the experimental farm (initial weight) and on d 21 and

42. Daily weight gain for each dietary treatment was calculated by difference between two initial and final weight of birds in each growth period (kg)/ number of days in the growth period. Feed consumption was recorded in the course of the whole starter and grower periods for each treatment, and the feed efficiency ratio were calculated subsequently.

#### **Carcass measurements**

At the end of the experiment, 12 birds per treatment were randomly taken to study carcass characteristics. Chicks were fasted for approximately 12 h, and then individually weighed, slaughtered, feathered and eviscerated. Weights of breast, thigh, heart, liver, spleen and gizzard were recorded. The percentage (% of live body weight) of carcass parts and organs was calculated.

#### Statistical analysis

Based a randomized complete design, all data were analyzed using the One-Way Anova procedure of SAS<sup>®</sup> (SAS, 1998) for analysis of variance. Significant differences among treatments were identified at 5% level by Duncan's multiple range tests (Duncan, 1955).

# **RESULTS AND DISCUSSION**

# **Growth performance**

The results of performance are presented in the Table 2. At the starter period of the experiment (1 to 21 days), birds supplemented with prebiotic and synbiotic had a greater (P<0.05) daily body weight gain (BWG) compared with control group. The daily BWG (from day 1 to 42) were increased (P<0.05) for birds supplemented with antibiotic and synbiotic compared with control treatment. The inclusion of all treatments, except probiotic, in the feed also promoted feed efficiency (1 to 21 and 1 to 42 days) as compared to the control group (p<0.05). Daily feed intake during the experimental period did not differ between treatments (p>0.05).

In general the positive effects of experimental additives on performance are in agreement with the results reported previously. Fairchild et al. (2001), who showed that flavomycin as antibiotic has favorable effects on the weight gain of broiler chickens. Esteve-garcia et al. (1997), observed that addition of flavomycin to a wheat-based ration could improve significantly the chickens feed efficiency ratio in all breeding periods (1 to 21 days and 22 to 42 days). In the present study, the beneficial effects of a probiotic, prebiotic and synbiotic products on broiler performance parameters including daily BWG and feed efficiency are in agreement with previous studies (Zulkifli et al., 2000; Pelicano et al., 2003; Cavit, 2004; Thitaram et al., 2005; Kermanshahi and Rostami, 2006; Navebpor et al., 2007; Falaki et al., 2010). In contrast, Gunal et al., (2006), Zhang et al., (2005), Jamroz et al., (2004) and Willis et al., (2007), reported that using these additives shed in the broiler ration had no significant effects on growth performance of broiler chickens. Variance among

	Starter	Grower (22-42 days)	
Ingredients —	(1-21 days)		
Corn	58.7	61	
Soybian meal	30	29	
Wheat bran	5	5	
Fish meal	2	0	
Soybian oil	1	2	
Oister shell meal	1.2	1	
DCP	1.07	1	
Vitamin and mineral perimix	0.5	0.5	
DL- Methionine	0.13	0.1	
L-lysine	0.15	0.25	
Salt	0.25	0.1	
Coccidiostat	0	0.05	
Total	100	100	
Nutrient content			
ME (Kcal/Kg)	2850	2950	
Crude protein (%)	20.48	18.44	
Crude fiber (%)	3.89	3.81	

 Table 1. Ingredient composition (as percent of dry matter) and calculated analysis of the basal diets.

Vitamin and mineral provided per kilogram of diet: vitamin A, 360000 IU; vitamin D3, 800000 IU; vitamin E, 7200 IU; vitamin K3, 800 mg; vitamin B1, 720 mg; vitamin B9, 400 mg; vitamin H2, 40 mg; vitamin B2, 2640 mg, vitamin B3, 4000 mg; vitamin B5, 12000 mg; vitamin B12, 6 mg; Choline chloraid, 200000 mg, Manganeze, 40000 mg, Iron, 20000 mg; Zinc, 40000 mg, coper, 4000mg; Iodine, 400 mg; Selenium, 80 mg.

	<b>G</b> .					
	Control	Antibiotic	Probiotic	Prebiotic	Synbiotic	
Daily BWG (g)						
1-21	24.35 <sup>b</sup>	25.45 <sup>ab</sup>	25.07 <sup>ab</sup>	26.55 <sup>a</sup>	26.70 <sup>a</sup>	
1-42	47.53 <sup>b</sup>	50.65 <sup>a</sup>	49.28 <sup>ab</sup>	49.47 <sup>ab</sup>	51.08 <sup>a</sup>	
Daily FI (g)						
1-21	49.89	48.72	49.00	49.68	49.68	
1-42	106.13	106.24	106.16	105.46	106.26	
Feed efficiency (g/g)						
1-21	0.48 <sup>b</sup>	0.52 <sup>a</sup>	0.51 <sup>ab</sup>	0.53 <sup>a</sup>	0.53 <sup>a</sup>	
1-42	0.44 <sup>b</sup>	0.47 <sup>a</sup>	0.46 <sup>ab</sup>	0.47 <sup>a</sup>	0.48 <sup>a</sup>	

Table 2. The main effects of treatments on growth performance of broiler chickens.

<sup>a,b</sup> means in each row with different superscripts are significantly different (p<0.05).

BWG= Body Weigh Gain, FI= Feed Intake.

reports of researchers could be related to differences in the type of probiotics and prebiotics, management and environmental conditions that exist in various experiments. It's suggested that under benefit management and/or environmental conditions, the effect of such feed additives may be worthless. On the other hand, the responses of breeding bird in a warm climate condition similar to the present environmental in south west of Iran

to these growth promoters may be better than an ideal condition. The results of some studies shown that growth stimulating antibiotics, increase the growth of broiler chickens by an increase in the uptake of nutrients (especially fatty acids and glucose), fixation of nitrogen and reduction in excretion of fat in the feces and microbial urea (Anderson et al., 1999). Also, antibiotics reduce the number of bacteria, toxins and their

	Control	Antibiotic	Probiotic	Prebiotic	Synbiotic
Breast	18.91 <sup>b</sup>	20.20 <sup>a</sup>	20.41 <sup>a</sup>	18.82 <sup>b</sup>	19.92 <sup>ab</sup>
Thigh	19.28 <sup>c</sup>	20.87 <sup>a</sup>	19.99 <sup>abc</sup>	19.59 <sup>bc</sup>	20.55 <sup>ab</sup>
Heart	0.45	0.46	0.45	0.41	0.41
Spleen	0.15	0.18	0.16	0.20	0.15
Liver	2.24	2.04	2.07	2.29	2.19
Gizzard	1.71 <sup>ab</sup>	1.63 <sup>ab</sup>	1.57 <sup>b</sup>	1.88 <sup>a</sup>	1.85 <sup>ab</sup>
Abdominal Fat pad	1.98 <sup>a</sup>	1.91 <sup>a</sup>	1.74 <sup>ab</sup>	1.62 <sup>b</sup>	1.86 <sup>ab</sup>

Table 3. The main effects of treatments on carcass characteristic (as percent of live body weight) of 42d broiler chickens.

<sup>a,b,c</sup> means in each row with different superscripts

secondary products in the GI tract (Gunal et al., 2006). Our findings showed that consumption of synbiotic, like that of antibiotics, had a positive effect on the body weight gain when compared to control treatment. The reason may be ascribed to the synergism of probiotic and prebiotic following the concurrent action of prebiotics fermentation by lactic acid bacteria in the GI tract and production of some acids by this group of bacteria, the pH of the GI tract is further reduced (Fuller, 1989). Reduction in pH is effective in controlling the population of pathogenic bacteria. During the infections due to pathogenic bacteria, lymphocytes crowd up to kill them, and after inflammation the thickness of the muscular layer increases (Gunal et al., 2006). It seems that in our study, due to the synergism between probiotic and prebiotic, followed by absorption of nutrients by the GI system, the birds under synbiotic treatment had the best feed efficiency in both start (1 to 21 days) and whole (1 to 42 days) of experimental periods.

# **Carcass composition**

The effects of dietary treatments on carcass characteristics and some internal organs of 42 days old broilers are shown in Table 3. The efficiency of breast was higher (p<0.05) in the antibiotic and probiotic treatments than in the control group, and it in the prebiotic and synbiotic groups did not differ significantly as compared to the control treatment. The antibiotic supplemented group had a greater (P<0.05) thigh percent compared with the control and prebiotic supplemented groups. However, the thigh percent did not show significant differences between the probiotic and prebiotic groups with the control treatment. Also, there was no significant difference in mean percentage yield of heart, spleen and liver between the fed additives treatments and control group. The birds under prebiotic treatment, had a higher gizzard percent as compared to the probiotic treatment, whereas the percent of abdominal fat pad was lower than in the antibiotic and control treatments (p<0.05).

The benefits of the antibiotic use on carcass

characteristics have been reported by other researchers (Woodward et al., 1988; Elwinger et al., 1998; Fidler et al., 2003). Kabir et al. (2004) have reported that adding 2 g probiotic per each liter of water consumed by broiler chickens, would increase the efficiency in their thigh and breast as compared with the control treatment. In Ammerman et al. (1989) study, adding 0.375% oligofructose to the bird's ration, on day 47, decreased the percent of abdominal fat. However, our findings on carcasse composition were in contrast to those of Pelicia et al. (2004); Pelicano et al. (2003); Willis et al. (2007) and Kannan et al. (2005). As pointed out before, these differences between reported results could be related to the mode of action of those feed additives is guite different, particularly their antimicrobial activity, the similar physiologic pattern was probably exerted by modifying intestinal pH, altering the composition and balance of intestinal flora, enhancing nutrient digestibility and improving growth rate and carcass characteristics. Also, by observing a reduction in the abdominal fat level of birds fed by prebiotic, it is suggested that this product may interfere in the accessibility to fat for formation of fat tissue in the birds.

# Conclusions

Supplementation of broiler diets with the feed growth promoters improved the growth responses compared with the unsupplemented control treatment. Also, based on obtaining better results for birds fed diets containing growth promoters (synbiotic), particularly in growth response, when compared to control treatment, it is concluded that by using non-antibiotic additives particularly mixing of both probiotic and prebiotic could obtained the advantages of antibiotic (performance) without their disadvantages.

# REFERENCES

Ammerman E, Quarles C, Twining PV (1989). Evaluation of

fructo-ligosaccharides on performance and carcass yield of male broilers. Poult. Sci., 68: 167.

- Anderson DB, Mc Cracken JJ, Aminov RI, Simpson JM, Mackie RI, Verstegen MWA, Gaskins HR (1999). Gut microbiology and growthpromoting antibiotics in swine. Pig News Inform., 20: 115N-122N.
- Bailey JS, Blankenship LC, Cox NA (1991). Effect of fructooligosaccharide on Salmonella colonization of the chicken intestine. Poult. Sci., 70: 2433-2438.
- Cavazzoni V, Adami A, Cstrivilli C (1998). Performance of broiler chickens supplemented with Bacillus coagulans as probiotic. Br. Poult. Sci., 39: 526-529.
- Cavit, A (2004). Effect of dietary probiotic supplementation on growth performance in the rock partridge (*Alectoris graeca*). Turk. J. Vet. Anim. Sci., 28: 887-891.
- Chichlowski M, Croom J, McBride BW, Daniel L, Davis G, Koci MD (2007a). Direct-fed Microbial primalac and salinomycin modulate whole-body and intestinal oxygen consumption and intestinal mucosal cytokine production in the broiler chick. Poult. Sci., 86: 1100-1106.
- Chichlowski M, Croom J, Edens FW, McBride BW, Qui R, Chiang CC, Daniel LR, Havenstein GB, Koci MD (2007b). Microarchitecture and spatial relationship between bacreria and ileal, cacal, and colonic epithelium in chicks fed a direct-fed microbial, primalac, and salinomycin. Poult. Sci., 86: 1121-1132.
- Duncan DB (1955). Multiple range test and multiple F-tests. Biometrics., 11: 1-42.
- Elwinger K, Berndtson E, Engstrom B, Fossum O, Waldenstedt L (1998). Effect of antibiotic growth promoters and anticoccidials on growth of clostridium prefringenes in the caeca and on performance of broiler chickens. Acta. Vet. Scand., 39: 433-441.
- Esteve-garcia E, Brufau J, Perez-vendrell A, Miquel A, Duven K (1997). Bioefficacy of enzyme preparations containing b-glucanase and xylanase activities in broiler diets based on barley or wheat, in combination with flavomycin. Poult. Sci., 76: 1728-1737.
- Fairchild AS, Grimes JI, Jones FT, Wineland MJ, Edens FW, Sefton AE (2001). Effects of Hen Age, Bio-Mos, and Flavomycin \_on Poult Susceptibility to Oral Escherichia coli Challenge. Poult. Sci., 80: 562-571.
- Falaki M, Shams Shargh M, Dastar B, Zrehdaran S (2010). Effects of different levels of probiotic and prebiotic on performance and carcass characteristics of broiler chickens. J. Anim. Vet. Adv., 9(18): 2390-2395.
- Fidler DJ, George B, Quarles CL, Kidd MT (2003). Broiler performance and carcass traits as affected by dietary liquid saccharopoly spora solubles concentrate. J. Appl. Poult. Res., 12: 153-159.
- Fuller R (1989). Probiotic in man and animal. J. Appl. Bacteriol., 66: 365-378.
- Gallaher DD, Khil J (1999). The effect of synbiotics on colon carcinogenesis in rats. J. Nutr., 129(7): 1483S-1487S.
- Gibson GR, Roberfroid MB (1995). Dietary modulation of human colonic microbiota: Introducing the concept of prebiotic. J. Nutr., 125: 1401-1412.
- Griggs JP, JacobJP (2005). Alternatives to Antibiotics for Organic Poultry Production. J. Appl. Poult. Res., 14: 750-756.
- Gunal M, Yayli G, Kaya O, Karahan N, Sulak O (2006). The effects of antibiotic growth promoter, probiotic or organic acid supplementation on performance, intestinal microflora and tissue of broiler. Int. J. Poult. Sci., 5: 149-155.

- Jamroz D, Wiliczkiewicz A, Orda J, Wertelecki T, Skorupin' ska J (2004). Response of broiler chickens to the diets supplemented with feeding antibiotic or mannan-oligosaccharides. Electr. J. Polish Agric. Univ., Ser., Anim. Husb., 7.
- Kabir SML, Rahman MM, Rahman MB, Rahman MM, Ahmed SU (2004). The dynamics of probiotics on growth performance and immune response in broilers. Int. J. Poult. Sci., 3: 361-364.
- Kannan M, Karunakaran R, Balakrishnan V, Prabhakar TG (2005). Influence of prebiotics supplementation on lipid profile of broilers. Int. J. Poult. Sci., 4(12): 994-997.
- Kermanshahi H, Rostami H (2006). Influence of supplemental dried whey on broiler performance and cecal flora. Int. J. Poult. Sci., 5: 538-543.
- Nayebpor M, Farhomand P, Hashemi A (2007). Effect of different levels of direct fed microbial (Primalac) on the growth performance and humoral immune response in broiler chickens. J. Anim. Vet. Adv., 6: 1308-1313.
- NRC (National Research Council), (1994). Nutrient Requirements of Poultry. 9th. Rev.(ed).National Academy Press, Washington, D. C.
- Patterson JA, Burkholder KM (2003). Application of prebiotics and probiotics in poultry production. Poult. Sci., 82: 627-631.
- Pelicano ERL, Souza PA, Souza HBA, Oba A, Norkus EA, Kodawara LM, Lima TMA (2003). Effect of different probiotics on broiler carcass and meat quality. Brazil J. Poult. Sci., 5: 207-214.
- Pelicia K, Mendes AA, Saldanha ESPB, Pizzolante CC, Takahashi SE, Moreira J, Garcia RG, Quinterio RR, Paz LCLA, Komiyama CM (2004). Use of prebiotics and probiotics of bacterial and yeast origin for freerange broiler chickens. Brazil J. Poult. Sci., 6: 163-169.
- Samli HE, Senkoylu N, Koc F, Kanter M, Agma A (2007). Effects of Enterococcus faecium and dried whey on broiler performance, gut histomorphology and microbiota. Arch. Anim. Nutr., 61: 42-49.
- Statistic Analysis System (1998). Painless windows, a handbook for SAS users. 2nd ed. Guelph (ON): Jodie Gilmore.
- Thitaram SN, Chung CH, Day DF, Hinton A, Bailey JS, Siragusa GR (2005). Isomaltooligosaccharide increases cecal bifidobacterium population in young broiler chickens. Poult. Sci., 84: 998-1003.
- Willis WL, Isikhuemhen OS, Ibrahim A (2007). Performance assessment of broiler chickens given mushroom extract alone or in combination with probiotic. Poult. Sci., 86: 1856-1860.
- Woodward SA, Hams RH, Miles RD, Janky DM, Ruiz N (1988). Research note: Influence of virginiamycin on yield of broilers fed four levels of energy. Poult. Sci., 67: 1222-1224.
- Wray C, Davies RH (2000). Competitive exclusion-An alternative to antibiotics. Vet. J., 59: 107-108.
- Zhang AW, Lee BD, Lee SK, Lee KW, An GH, Song KB, Lee CH (2005). Effects of Yeast (Saccharomyces cerevisiae) Cell Components on Growth Performance, Meat Quality, and Ileal Mucosa Development of Broiler Chicks. Poult. Sci., 84: 1015-1021.
- Zulkifli I, Abdullah N, Azrin NM, Ho YW (2000). Growth performance and immune response of two commercial broiler strains fed diets containing Lactobacillus cultures and oxytetracycline under heat stress conditions. Br Poult Sci., 41: 593-597.