

Short Communication

Pathogenic diversity of *Sclerotium rolfsii* isolates, a potential biocontrol agent against *Parthenium hysterophorus* L.

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Accepted 16 March, 2011

Parthenium hysterophorus L. is a wide-spread weed creating problems for agriculture and public health. Microbes and their by products is now proved to be a worthy alternative to toxic chemicals used for weed management. We determined the relative pathogenicity of ten *Sclerotium rolfsii* isolates recovered from diseased parts of *Parthenium* collected during a survey of various habitats in Central India. There was a considerable diversity amongst various isolates. The isolate designated Par # 02 from Jabalpur showed the maximum disease incidence (80%) whereas isolate Par#10 also from Jabalpur showed the lowest infection rate against targeted hosts (30%). The wide range of pathogenicity among *S. rolfsii* support the view that strain breeding for biological control of *P. hysterophorus* L. is warranted.

Key words: *Parthenium hysterophorus*, variability, pathogenicity, *Sclerotium rolfsii*.

INTRODUCTION

Parthenium (*Parthenium hysterophorus* L.) is a cosmopolitan weed. In India it is responsible for several severe health problems viz. dermatitis, asthma, bronchitis etc in humans and animals (Pandey et al., 1996). The weed is also imposing serious effect to economically important crops and forest plants. Ever since the weed became a menace in various parts of world including India, efforts are being made to manage the weed employing different methods. However, so far no single method has been proven satisfactory as each method suffers from one or more limitations (Templeton 1990, Hasija et al., 1994). Exploitation of microorganisms especially plant pathogenic fungi are now emerging as an effective and eco-friendly alternative to conventional methods of weed control (Pandey et al., 2003; 2004). *Sclerotium rolfsii* Sacc (teleomorph: *Athelia rolfsii*) Curzi Tu Kimbrough is a potential mycoherbicidal agent, incite severe collar rot disease in *Parthenium* (Mishra et al., 1996 a, b; Pandey et al., 1998; Shukla and Pandey 2006).

Although, the pathogen is responsible for severe damage to the weed but wide host range of the species creates

doubt about its suitability as mycoherbicides. However, there are several reports where various isolates of *S. rolfsii* have showed significant variations not only in their morphology but also in their pathological behavior (Harlton et al., 1995; Nalim et al., 1995; Okabe et al., 1998, 2000; Sharma et al., 2002; Shukla and Pandey, 2007). Therefore, present investigation was carried out to study the pathogenic variability and justify the separate identity of the *S. rolfsii* isolates associated with *Parthenium*.

MATERIALS AND METHODS

Test isolates and their maintenance

Ten isolates of *S. rolfsii* incites collar rot disease were recovered from *Parthenium* from diverse geographic locations. These were maintained on Potato Dextrose Agar (Potato-200 g; Dextrose-20 g; Agar-18 g; D/W 1000 ml) in Petri dishes or in test tube slants at room temperature (Agrawal and Hasija, 1986).

Preparation of inoculum

A granular formulation of the inoculum was used for the study. Plastic bags containing 25 g of wheat (pre boiled) sterilized at 121°C and 15 lb pressure for 15 min was seeded with 5 mm² disc of all the 10 *S. rolfsii* isolates obtained from 7 d old cultures grown in PDA Plates. Inoculated bags were kept in incubator (Remi) at 28 ± 2°C for 15 - 20 days (till

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Table 1. Origin and Virulence of *S. rolfsii* isolates after 10 days of inoculation.

S/No.	Isolates	Place of isolation	%Disease incidence
1	Par#01	Jabalpur	45
2	Par#02	Jabalpur	80
3	Par#03	Seoni	56.6
4	Par#04	Singrauli	47
5	Par#05	Sidhi	52
6	Par#06	Kareli	62
7	Par#07	Mandla	55
8	Par#08	Jabalpur	73
9	Par#09	Narsingpur	47
10	Par#10	Jabalpur	30
11.	Control		No effect
	SEM		± 2.1
	C.D		±5.1

Amount of Inoculum: 10sclerotia/pot; No of Pots used: 3pots/treatment. Age of plant: 2 weeks old; R.H = 85-90%. Values given in the table are tested by F test and are significant a 5% level of significance

sclerotia formation) (Moctezuma et al., 2006).

Evaluation of aggressiveness of *S.rolfsii* isolates

Parthenium seeds were sown at 5 seeds/pot in eathern pots (10 cm) containing sterilized soil sand and peat (1:1:1). Two week after germination (4 leaf stage) these were inoculated with 10 dried sclerotia of each *S. rolfsii* isolate. A non inoculated pot served as control. Sclerotia were placed beneath the surface of the soil, contacting the stem of the plant. All the plants were maintained in the green house and % disease incidence was recorded after 2nd day to 10th of inoculation by the following formula and the data were analyzed by analysis of variance (Moctezuma et al., 2006).

RESULT AND DISCUSSION

Four distinct pathogenicity reactions were observed for all the 10 isolates tested against *Parthenium* (Table 1). Pathogenicity reactions ranged from 30 to 80%. Disease reactions varied with the isolates which ranged from yellowing of entire plant and formation of sclerotia to wilting and finally death of plant (highly aggressive). As depicted in Table 1 isolate Par# 02 exhibited maximum disease incidence followed by Par#08 isolate. Rest of the isolates also showed more than 50% disease incidence except Par#10 where only 30% disease incidence was recorded. Significant variation in pathogenicity and virulence of various patho-gens at different levels viz., generic, species levels or even at intraspecific level have also recorded by many workers (Agnihotri and Ansari, 2000). Variation in virulence has been correlated with synthesis of oxalic acid and of enzymes (Punja et al., 1985; Agnihotri and Ansari, 2000).

Therefore, significant variation existed between all the isolates tested in the present investigation. Par#02 isolate exhibited maximum disease incidence, has immense poten-

to be develop as mycoherbicide agent against *P. hystero-phorus*.

ACKNOWLEDGEMENT

We are grateful to Head, Department of Biological Sciences, R.D. University, Jabalpur for providing necessary laboratory facilities. We are also thankful to Madhya Pradesh Council of Science and Technology, Bhopal, Council of Scientific and Industrial Research, New Delhi and DBT, New Delhi for financial supports.

REFERENCES

- Agrawal GP, Hasija SK (1986). Microorganisms in the laboratory: A laboratory guide for Mycology, Microbiology and Plant Pathology. Print house India, Lucknow. p.155.
- Ansari MM, Agnihotri SK (2000). Morphological, physiological and pathological variation among *Sclerotium rolfsii* isolates of Soybean. Indian Phytopathol. 53(1): 65-67.
- Harlton CE, Levesque CA, Punja ZK (1995). Genetic diversity in *Sclerotium (Athelia) rolfsii* and related species. Phytopathol. 85:1269-1281.
- Hasija SK, Rajak RC, Pandey AK (1994). Microbes in the management of obnoxious weed, In: Vistas in seed Biology (Eds. Singh, T and P.C. Trivedi) Vol. I Print Well Jaipur, pp. 82 – 104.
- Mishra J, Pandey AK, Hasija SK (1996a). Mycoherbicidal potential of *Sclerotium rolfsii* Sacc. against *Parthenium*: Factors affecting in vitro growth and sclerotial formation. J. Pathol. Res. 9(1): 19 – 24.
- Mishra J, Pandey AK, Hasija SK (1996b). Mycoherbicidal potential of *Sclerotium rolfsii* Sacc. against *Parthenium hysterophorus* L.: histopathological studies. Indian J.Appl.Pure Biol.11:73-77.
- Moctezuma- Flores HK, Motes- Belmon R, Jimenz-Perez A, Nava-Juarez R (2006). Pathogenic diversity of *Sclerotium rolfsii* isolates from Mexico and potential control of southern blight through solarization and organic amendments. Crop Protec.25:195-201.
- Nalim FA, Starr JL, Woodward KG, Segner S, Keller NP (1995). Mycelial compatibility groups in Texas Peanut field populations of *Sclerotium rolfsii*. Phytopathol. 85: 1507-1512.
- Okabe I, Morikawa C, Matsumoto N, Yokoyama K (1998). Variation in *Sclerotium rolfsii* isolates in Japan. Mycosci. 39: 399-407.
- Okabe I, Morikawa C, Matsumoto N (2000).Variation in southern blight fungus in Japan detected by ITS-RFLP analysis. Japan Agri. Res. Quat. 34:93-97.
- Pandey AK, Mishra J, Rajak RC, Hasija SK (1996). Potential of indigenous strains of *Sclerotium rolfsii* Sacc. for the management of *Parthenium hysterophorus* L.: A serious threat to biodiversity in India. In: Herbal Medicines, Biodiversity and Conservation Strategies (Eds. Rajak, R C and Rai M K). International Book Distributors, Dehradun. pp.104-138.
- Pandey AK, Mishra Jyoti, Hasija SK (1998). Effect of inoculum on mycoherbicidal potential of *Sclerotium rolfsii* against *Parthenium*. J.Mycol.PI.Pathol.28: 284-287.
- Pandey AK, Singh Jaya, Shrivastava GM, Rajak RC (2003). Fungi as herbicides: Current status and future prospects. In: Plant Protection: Biological Approach (Eds. Trivedi P.C.), Aavishkar Publishers, Distributors Jaipur, India: 305-339
- Pandey AK, Pandey Archana,Shrivastava GM, Rajak RC (2004). Potential of microorganisms for the management of *Lantana camara* in India: Possibilities and Prospects. In: Microbiology and biotechnology for Sustainable Development (Ed. P.C. Jain).CBS Publishers and Distributors, New Delhi. pp.42-58.
- Punja ZK, Smith VL, Campbell CL, Jenkins SF (1985).Sampling and extraction procedure to estimate numbers, spatial pattern and temporal distribution of sclerotia of *Sclerotium rolfsii* in soil. Plant Dis. 69: 469-474.
- Sharma B K, Singh U P, Singh K P (2002). Variability in Indian isolates of *Sclerotium rolfsii*. Mycologia 94(6): 1051-1058.

Shukla Rekha, Pandey AK (2006). Maximization of production of oxalic acid from *Sclerotium rolfsii*, a mycoherbicidal agent against Parthenium. Ann. Pl. Protect. Sci. 14(1): 202-205.

Shukla Rekha, Pandey A K (2007). Diversity in mycoherbicidal agent *Sclerotium rolfsii* isolates from Central India. J.Mycol. Pl. Pathol. 37(3): 514-518.

Templeton GE (1990). Weed control with pathogens: Future needs and directions: In: Microbes and microbial products as herbicides (Hoagland R.E Eds) ACS Symposium series 439 American Chemical Society, Washington DC: pp. 320-29.